An Asymmetric Synthesis of D-1,6-Diepicastanospermine

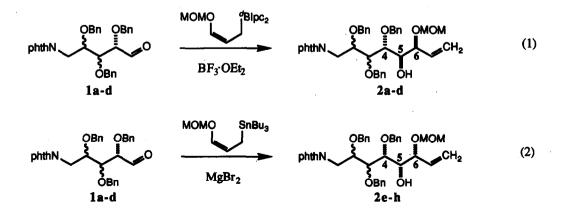
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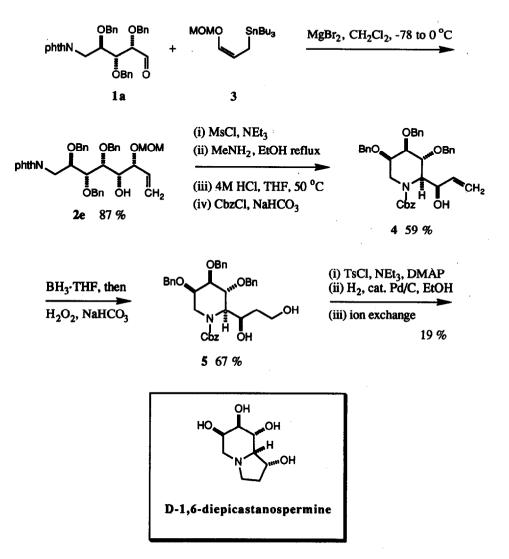
Key Words: castanospermine, 1,6,7,8-tetrahydroxyindolizidine, glucosidase inhibitor.

Abstract: D-1,6-Diepicastanospermine has been prepared via reaction of an allylstannane with the arabinose derivative 1a. This reaction was almost 100 % stereoselective, but the corresponding allylations of aldehydes 1b - d gave mixtures of isomers. Variable stereodifferentiation in these reactions can be attributed to the influence of the distal (γ) chiral center.

Castanospermine and its stereoisomers are worthwhile synthetic targets due to their proven¹⁻¹² and potential biological activities, particularly with respect to chemotherapy of AIDS, cancer, and diabetes. Nevertheless, most of the thirty-one stereoisomers of castanospermine have yet to be prepared or isolated, despite the efforts of several research groups.¹³ Our response to obtaining samples for testing has been to design syntheses that can be readily adapted to give several stereoisomeric products without significant changes in the experimental protocol used for most of the steps. So far, we have shown eight castanospermine stereoisomers can, in principle, be prepared using the reaction shown in equation 1.¹⁴ The allylation methodology used in this transformation¹⁵ is confined to preparations of intermediates with the 4,5-*anti*, 5,6-*syn* relative stereochemistry, *eg* **2a-d**, as shown. This paper describes efforts to prepare the 4,5-*syn*, 5,6-*syn* isomers **2e-h**, and an asymmetric synthesis of D-1,6-diepicastanospermine.

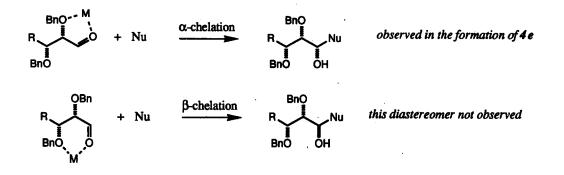


Anti stereochemistry about the 4,5-junction in compounds **2a-d** (eq. 1) corresponds to allylation of the substrates 1 in a Felkin-Anh sense.¹⁶ We sought to overcome this preference using a chelation controlled,¹⁷ syn selective attack with an allylstannane reagent (eq. 2).¹⁸⁻²⁰ Fortunately, this worked very well for the arabinose derivative **1a**; only one diastereomer of the product was detected by ¹H/¹³C NMR, and the HPLC trace of the crude reaction mixture was relatively clean.



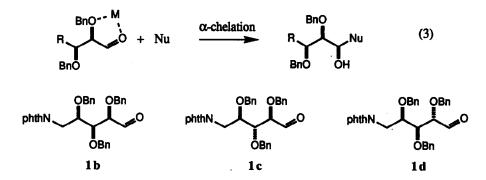
Assignment of stereochemistries for products 2 is non-trivial, especially when they are formed via a reaction which generates two asymmetric centers simultaneously. In the event, stereochemical characterization of compound 2e was performed by completing the synthesis; the product obtained was spectroscopically identical to a sample of D-1,6-diepicastanospermine prepared via another route.^{21,22} Apart

from the allylation reaction, each step in this synthesis is very similar to the analogous transformations described in our first paper on syntheses of castanospermine stereoisomers.^{13,23} Substrates 2 are capable of coordinating to MgBr₂ via the α , β , and possibly, the γ -benzyloxy groups; one might anticipate coordination to the α and β -groups to be the most significant. In terms of substrate-controlled diastereoselectivity, the α and β -asymmetric centers are mismatched for chelation controlled processes (as illustrated below).



(R = chiral group)

The allylation of 1a in our synthesis of 1,6-diepicastanospermine is consistent with attack on an α benzyloxy-chelated intermediate, and not with coordination of the β -benzyloxy substituent. This oversimplistic model implies the stereochemistry at the β -center is irrelevant and allylations of substrates with other relative α/β -configurations should also react with good selectivity, as depicted in equation (3).



In practice, however, allylations of aldehydes 1b-d with allylstannane 3 gave a mixture of diastereomers in each case (1b, 79:13:8:<1; 1c, 77:23:<1:<1; 1d, 90:10:<1:<1; stereochemistries not assigned). Qualitative differences in reaction rates were also observed, the arabinose derivative 1a being the most reactive.

It would be extremely inconvenient to prepare castanospermine stereoisomers via allylations of aldehydes 1b-d. This work does, however, provide an excellent illustration of the potential influence of distal chiral centers in substrate-controlled diastereoselectivity. This type of phenomenon has been reported for other reaction types,^{24,25} and more will undoubtedly emerge as the field of asymmetric synthesis expands.

Acknowledgments

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- 1. R. Saul, R. J. Molyneux, and A. D. Elbein, Arch. Biochem. Biophys., 1984, 230, 668.
- Y. T. Pan, H. Hori, R. Saul, B. A. Sanford, R. J. Molyneux, and A. D. Elbein, *Biochemistry*, 1983, 22, 3975.
- 3. R. Saul, J. P. Chambers, R. J. Molyneux, and A. D. Elbein, Arch. Biochem. Biophys., 1983, 221, 593.
- 4. R. Saul, R. J. Molyneux, and A. D. Elbein, Arch. Biochem. Biophys., 1984, 230, 668.
- 5. R. Saul, J. J. Ghidoni, R. J. Molyneux, and A. D. Elbein, Proc. Natl. Acad. Sci. U. S. A., 1985, 82, 93.
- 6. V. W. Sasak, J. M. Ordovas, A. D. Elbein, and R. W. Berninger, Biochem. J., 1985, 232, 759.
- 7. T. Szumilo, G. P. Kaushal, and A. D. Elbein, Arch. Biochem. Biophys., 1986, 247, 261.
- 8. B. C. Campbell, R. J. Molyneux, and K. C. Jones, J. Chem. Ecol., 1987, 13, 1759.
- 9. H. Hori, Y. T. Pan, R. J. Molyneux, and A. D. Elbein, Arch. Biochem. Biophys., 1984, 228, 525.
- I. C. d. Bello, D. Mann, R. J. Nash, and B. Winchester. "Castanospermine and other Glycosidase Inhibitors." Lipid Storage Disorders. Salvayre, Douste-Blazy and Gatt ed. 1988 Plenum. New York.
- 11. A. D. Elbein, CRC Critical Reviews in Biochemistry, 1984, 16, 21.
- G. W. J. Fleet, A. Karpas, R. A. Dwek, L. E. Fellows, A. S. Tyms, S. Petursson, S. K. Namgoong, N. G. Ramsden, P. W. Smith, J. C. Son, F. Wilson, D. R. Witty, G. S. Jacob, and T. W. Rademacher, *Febs Lett*, 1988, 237, 128.
- 13. K. Burgess, and I. Henderson, Tetrahedron, 1992, in the press.
- 14. K. Burgess, D. A. Chaplin, I. Henderson, Y. T. Pan, and A. D. Elbein, J. Org. Chem., 1992, 57, 1103.
- 15. H. C. Brown, P. K. Jadhav, and K. S. Bhat, J. Am. Chem. Soc., 1988, 110, 1535.
- K. N. Houk, M. N. Paddon-Row, N. G. Rondan, Y. Wu, F. K. Brown, D. C. Spellmeyer, J. T. Metz, Y. Li, and R. J. Loncharich, *Science*, 1986, 231, 1108.
- 17. M. T. Reetz, Angew. Chem. Int. Ed. Eng., 1984, 23, 556.
- 18. G. E. Keck, D. E. Abbott, and M. R. Wiley, Tetrahedron Lett., 1987, 28, 139.
- 19. M. Korceda, and Y. Tanaka, Tetrahedron Lett., 1987, 28, 143.
- 20. Y. Yamamoto, and S. Nishi, J. Org. Chem., 1988, 53, 3597.
- G. W. J. Fleet, N. G. Ramsden, R. J. Nash, L. E. Fellows, G. S. Jacob, R. J. Molyneux, I. C. d. Bello, and B. Winchester, *Carbohydrate Res.*, 1990, 205, 269.
- We thank Dr Gearoge Fleet (Oxford) for kindly providing NMR spectra of an authentic sample of 1,6diepicastanospermine.
- 23. D. L. Comins, and J. D. Brown, Tetrahedron Lett., 1986, 27, 4549.
- 24. Y. Yamamoto, H. Nemoto, R. Kikuchi, H. Komatsu, and I. Suzuki, J. Am. Chem. Soc., 1990, 112, 8598.
- 25. W. R. Roush, J. Org. Chem., 1991, 56, 4151.